

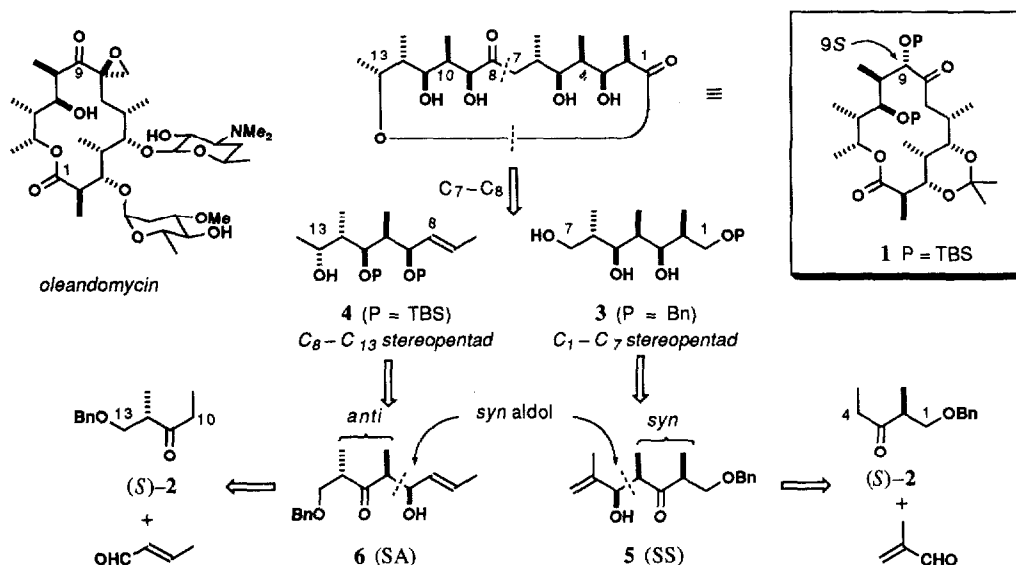
## Studies in Macrolide Synthesis: A Stereocontrolled Synthesis of a (9*S*)-Macrolide Intermediate for Oleandomycin Using Chiral Boron Reagents.

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**Abstract:** The (9*S*)-macrolide **1** (P = TBS) was prepared in 14 steps (5% yield) with 63% overall ds starting from the ethyl ketone (*S*)-**2**. The C<sub>1</sub>-C<sub>7</sub> and C<sub>8</sub>-C<sub>13</sub> segments, **3** and **4**, were obtained *via* boron enolate aldol reactions mediated by (+)- and (-)-(Ipc)<sub>2</sub>BOTf, respectively.

Aldol reactions of chiral boron enolates are useful for the enantio- and diastereoselective synthesis of the characteristic polypropionate segments of macrolide antibiotics.<sup>1</sup> Until now, this approach has largely relied upon the use of a suitable chiral auxiliary attached to a propionate enolate or equivalent.<sup>1,2</sup> We have recently adopted an alternative strategy using the easily prepared chiral boron reagents, (+)- and (-)-disopinocampheylboron triflate, to control enolization stereoselectivity and the enolate  $\pi$ -face selectivity in aldol reactions of both achiral<sup>3</sup> and chiral<sup>4</sup> ethyl ketones with aldehydes. We now report an application of this methodology in the synthesis of the 14-membered macrolide **1** (P = TBS), an intermediate in a projected synthesis of the clinically-important antibiotic oleandomycin.<sup>5,6</sup> In this synthesis (**Scheme 1**), six of the ten stereogenic centres in **1** are controlled by two reagent-controlled aldol reactions of the *same* chiral ethyl ketone (*S*)-**2**.



Scheme 1

**Scheme 1** outlines our retrosynthetic analysis of the (9*S*)-macrolide **1**, involving disconnection to the C<sub>1</sub>-C<sub>7</sub> and C<sub>8</sub>-C<sub>13</sub> stereopentad segments **3** and **4**. These should be available in turn from the  $\beta$ -hydroxyketones **5** and **6**, both of which can be obtained<sup>4b</sup> by *syn*-selective aldol reactions of the ethyl ketone (*S*)-**2** using reagent control from (+)- and (-)-diisopinocampheylboron triflate ((Ipc)<sub>2</sub>BOTf), respectively.

The synthesis of the (9*S*)-macrolide **1**, starting from the ethyl ketone (*S*)-**2** (97% ee),<sup>7</sup> is shown in **Scheme 2** and outlined below.

The *syn-syn* aldol isomer **5** (SS),<sup>4b,8</sup> required for the C<sub>1</sub>–C<sub>7</sub> segment **3**, was prepared in enantiomerically pure form *via* (+)-(Ipc)<sub>2</sub>BOTf mediated enolisation of (*S*)-**2** giving the corresponding *Z* enol diisopinocampheylborinate **7**, followed by addition of methacrolein. Similarly, the *syn-anti* aldol isomer **6** (SA),<sup>4b</sup> required for the C<sub>8</sub>–C<sub>13</sub> segment **4**, was prepared *via* (–)-(Ipc)<sub>2</sub>BOTf mediated enolisation of (*S*)-**2** giving the *Z* enol diisopinocampheylborinate **8**, followed in this case by addition of crotonaldehyde. This led to 90% diastereoselectivity for **7** → **5**, [ $\alpha$ ]<sub>D</sub><sup>20</sup> = +46.3° (*c* 4.0, CHCl<sub>3</sub>), and 89% diastereoselectivity for **8** → **6**, [ $\alpha$ ]<sub>D</sub><sup>20</sup> = +27.2° (*c* 6.3, CHCl<sub>3</sub>), where chromatography on silica gel allowed separation from the minor *syn* isomers.

For a *rapid* synthesis of the C<sub>1</sub>–C<sub>7</sub> segment **3** from the methacrolein aldol adduct **5**, we desired stereoselective alkene hydroboration and ketone reduction to set up the stereogenic centres at C<sub>3</sub> and C<sub>6</sub>. We anticipated from related studies<sup>9</sup> that this should be possible in a one-pot reaction of **5** with the sterically demanding borane (+)-(Ipc)<sub>2</sub>BH.<sup>2b,10</sup> Thus, treatment of **5** with (+)-(Ipc)<sub>2</sub>BH (3 equiv, Et<sub>2</sub>O, 2 h), followed by oxidative workup (*m*CPBA), gave only *two* out of the four possible triols by HPLC analysis. These were the desired triol **3**,<sup>8</sup> [ $\alpha$ ]<sub>D</sub><sup>20</sup> = +9.3° (*c* 2.3, CHCl<sub>3</sub>), and the epimeric triol at C<sub>6</sub>, which were obtained in a ratio of 92:8 and in 68% yield. The five contiguous stereogenic centres spanning C<sub>2</sub>–C<sub>6</sub> in oleandomycin were thus efficiently set up in only *two* steps from (*S*)-**2**.

The synthesis of the C<sub>8</sub>–C<sub>13</sub> segment **4** required a selective reduction of the ketone in the crotonaldehyde aldol adduct **6**, *i.e.* **6** → **9**, 11-*syn* diol **9**. This was accomplished with ≥97% ds (single isomer by 250 MHz <sup>1</sup>H NMR) and 79% yield by LiBH<sub>4</sub> reduction of the dibutylboron aldolate derived from reaction of **6** with <sup>n</sup>Bu<sub>2</sub>BOMe.<sup>11</sup> Next, a three-step sequence of (i) bis-TBS protection with <sup>t</sup>BuMe<sub>2</sub>SiOTf, (ii) debenzoylation using lithium di-*tert*-butylbiphenyl<sup>12</sup> in THF, and (iii) Swern oxidation (warming from –78 °C only as far as –23 °C after addition of Et<sub>3</sub>N, to avoid β-elimination), provided the aldehyde **10** in readiness for stereoselective methyl addition at C<sub>13</sub>. We required the Felkin-Cram adduct, and, of a number of reagents screened, MeMgCl gave both the highest yield and highest stereoselectivity. Thus, addition of MeMgCl to **9** at low temperature (–100 °C) gave the desired (13*R*) alcohol **4**,<sup>8</sup> [ $\alpha$ ]<sub>D</sub><sup>20</sup> = +5.3° (*c* 3.6, CHCl<sub>3</sub>), with 88% diastereoselectivity and in 73% yield from **9**. This completed the construction of the five stereogenic centres spanning C<sub>9</sub>–C<sub>13</sub> in six steps from (*S*)-**2**.

Protection of the C<sub>13</sub> hydroxyl in **4** as the BOM ether and subsequent ozonolysis gave the aldehyde **11** (92%), [ $\alpha$ ]<sub>D</sub><sup>20</sup> = –1.9° (*c* 9.0, CHCl<sub>3</sub>), in preparation for coupling with a nucleophilic C<sub>1</sub>–C<sub>7</sub> fragment derived from **3**. Based on our previous studies,<sup>5a</sup> we chose to convert **3** into the known phenyl sulphoxides **12**. This was achieved by a straightforward five-step sequence in 73% overall yield: (i) selective tosylation of the primary hydroxyl; (ii) acetonide formation; (iii) hydrogenolysis of the benzyl ether; (iv) displacement of the tosylate by LiSPh; (v) NaIO<sub>4</sub> oxidation to the sulphoxides **12** (identical in all respects with that made previously<sup>5a</sup>).

Coupling of the C<sub>1</sub>–C<sub>7</sub> and C<sub>8</sub>–C<sub>13</sub> segments was achieved by lithiation of **12** (LDA, 2.2 equiv, DME, –78 °C) followed by addition of the aldehyde **11**. This led to a complex mixture of adducts **13** in 84% yield, which were not separated.<sup>13</sup> Treatment of the adducts **13** with W–2 Raney nickel was followed by Swern oxidation and further oxidation of the C<sub>1</sub> aldehyde to the acid **14** using buffered KMnO<sub>4</sub>.<sup>14</sup> Deprotection of the BOM ether at C<sub>13</sub> was then achieved by hydrogenolysis to give the required *seco*-acid **15** (34% overall from **13**). This *seco*-acid was successfully macrolactonised in good yield (60%) under standard Yamaguchi conditions<sup>15</sup> to give the 14-membered macrolide, **15** → **1**,<sup>8</sup> [ $\alpha$ ]<sub>D</sub><sup>20</sup> = –3.8° (*c* 0.3, CHCl<sub>3</sub>).

In summary, we have achieved a short and highly stereocontrolled synthesis of an advanced macrolide intermediate for oleandomycin (14 steps from (*S*)-**2** *via* **6** in 5% yield; 63% overall ds). This relies on two *syn*-selective aldol reactions under reagent control, **2** → **5** and **2** → **6**, and a novel one-pot reduction/hydroboration, **5** → **3**. This work demonstrates that efficient synthesis of macrolides can be achieved using aldol reactions controlled by chiral boron reagents. Studies towards the elaboration of **1** into oleandomycin are underway.



## References and Notes

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- (S)-**2** was prepared from (S)-(+)-methyl 3-hydroxy-2-methylpropionate (Aldrich), in three steps and 62% yield (not from the (R)-(-)-propionate as we erroneously reported earlier), see ref 4b and 4e.
- All new compounds gave spectroscopic data in agreement with the assigned structures. **3** had  $^1\text{H}$  NMR  $\delta$  ( $\text{CDCl}_3$ , 400 MHz) 7.37-7.26 (5H, m), 4.49 (2H, ABq,  $J = 12.0$  Hz), 3.80 (1H, dd,  $J = 6.5, 3.1$  Hz), 3.70 (1H, dd,  $J = 9.4, 1.8$  Hz), 3.70 (1H, dd,  $J = 10.7, 3.9$  Hz), 3.65 (1H, dd,  $J = 10.7, 8.3$  Hz), 3.46 (1H, dd,  $J = 9.2, 4.7$  Hz), 3.43 (1H, dd,  $J = 9.2, 5.0$  Hz), 1.98 (1H, qddd,  $J = 6.9, 6.5, 5.0, 4.7$  Hz), 1.88 (1H, ddqd,  $J = 9.4, 8.3, 6.9, 3.9$  Hz), 1.80 (1H, qdd,  $J = 7.0, 3.1, 1.8$  Hz), 1.07 (3H, d,  $J = 6.9$  Hz), 0.96 (3H, d,  $J = 7.0$  Hz), 0.72 (3H, d,  $J = 6.9$  Hz);  $^{13}\text{C}$  NMR  $\delta$  ( $\text{CDCl}_3$ , 100 MHz) 138.1, 128.4, 127.6, 127.5, 82.6, 79.4, 74.1, 73.3, 69.1, 37.2, 36.8, 36.3, 13.4, 13.2, 5.5; HRMS (CI,  $\text{NH}_3$ )  $[\text{M}+\text{H}]^+$  found 297.2071,  $\text{C}_{17}\text{H}_{29}\text{O}_4$  requires 297.2066; **4** had  $^1\text{H}$  NMR  $\delta$  ( $\text{CDCl}_3$ , 400 MHz) 5.51 (1H, dq,  $J = 15.5, 6.0$  Hz), 5.38 (1H, ddq,  $J = 15.5, 6.8, 1.5$  Hz), 4.27 (1H, qd,  $J = 6.4, 1.8$  Hz), 3.96 (1H, dd,  $J = 6.8, 5.0$  Hz), 3.72 (1H, dd,  $J = 6.7, 2.0$  Hz), 3.50 (1H, br s), 1.84 (1H, qdd,  $J = 7.1, 6.7, 5.0$  Hz), 1.68 (3H, br d,  $J = 6.0$  Hz), 1.59 (1H, qdd,  $J = 7.1, 2.0, 1.8$  Hz), 1.11 (3H, d,  $J = 6.4$  Hz), 0.99 (3H, d,  $J = 7.1$  Hz), 0.95 (3H, d,  $J = 7.1$  Hz), 0.90 (9H, s), 0.86 (9H, s), 0.09 (3H, s), 0.08 (3H, s), -0.01 (3H, s), -0.03 (3H, s);  $^{13}\text{C}$  NMR  $\delta$  ( $\text{CDCl}_3$ , 100 MHz) 133.2, 126.6, 79.3, 75.8, 66.6, 43.7, 41.2, 26.2, 25.9, 20.9, 18.4, 18.2, 17.6, 12.1, 11.4, -3.5, -3.6, -3.8, -4.9; HRMS (CI,  $\text{NH}_3$ )  $[\text{M}+\text{H}]^+$  found 431.3375,  $\text{C}_{23}\text{H}_{51}\text{O}_5\text{Si}_2$  requires 431.3377; **1** had  $^1\text{H}$  NMR  $\delta$  ( $\text{CDCl}_3$ , 400 MHz) 5.19 (1H, q,  $J = 6.4$  Hz), 4.47 (1H, br d,  $J = 4.4$  Hz), 3.82 (1H, d,  $J = 10.6$  Hz), 3.56 (1H, br d,  $J = 10.8$  Hz), 3.14 (1H, d,  $J = 8.7$  Hz), 3.00 (1H, m), 2.65 (1H, dq,  $J = 10.8, 6.6$  Hz), 2.54-2.40 (2H, m), 1.96 (1H, dq,  $J = 10.6, 6.6$  Hz), 1.60-1.50 (2H, m), 1.41 (3H, s), 1.40 (3H, s), 1.22 (3H, d,  $J = 6.4$  Hz), 1.08 (3H, d,  $J = 6.6$  Hz), 1.02 (3H, d,  $J = 6.7$  Hz), 0.97 (3H, d,  $J = 6.7$  Hz), 0.94 (3H, d,  $J = 6.6$  Hz), 0.91 (3H, d,  $J = 6.6$  Hz), 0.90 (9H, s), 0.89 (9H, s), 0.10 (3H, s), 0.06 (3H, s), 0.01 (3H, s), -0.03 (3H, s);  $^{13}\text{C}$  NMR  $\delta$  ( $\text{CDCl}_3$ , 100 MHz) 212.5, 173.8, 99.7, 81.7, 75.6, 72.0, 71.4, 71.4, 43.7, 40.2, 40.1, 38.6, 32.8, 30.5, 29.7, 26.3, 25.7, 19.5, 18.8, 18.4, 18.0, 15.6, 12.4, 10.1, 8.7, 7.7, -2.8, -4.9, -5.0, -5.7; HRMS (CI,  $\text{NH}_3$ )  $[\text{M}+\text{NH}_4]^+$  found 660.4692,  $\text{C}_{34}\text{H}_{70}\text{NO}_7\text{Si}_2$  requires 660.4690.
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